Effects of Red light on the Growth of *Zantedeschia* Plantlets in vitro and Tuber Formation Using Light-emitting Diodes

Ruey-Chi Jao¹

Department of Information Management, Chienkuo Technology University

Chien-Chou Lai

Department of Bio-Industrial Mechatronics Engineering, National Taiwan University

Wei Fang and Sen-Fuh Chang

Department of Bio-Industrial Mechatronics Engineering, National Taiwan University

Additional index words. LED, Zantedeschia, AC power, red light

Abstract. Effects of light generated by red and blue light-emitting diodes on the photomixotrophic growth of Zantedeschia jucunda 'Black Magic' plantlets in vitro and tuber formation after transplant under the same PPF and photoperiod were investigated. All five treatments had the same photosynthetic photon flux (PPF, $80 \pm 5 \mu mol \cdot m^{-2} \cdot s^{-1}$) and photoperiod (16 hours daytime/8 hours nighttime), leading to the same daily light integral. Results showed that the tubular fluorescent lamp (TFL) treatment had the highest value on chlorophyll content and dry weight accumulation than other treatments using light-emitting diodes (LEDs). In LED treatments, there were no significant differences on dry weight and growth rate but with significant differences on chlorophyll content and plant height when blue light LEDs were added. It revealed that blue light was involved in plant height and chlorophyll development control mechanism. Results also showed that minor difference in 28 days of transplant production (in vitro) among treatments does not extends after 6 months of tuber formation stage grown in greenhouse. At present, blue LEDs cost much more than that of red LEDs, results of this study suggested that using red LEDs alone and powered with AC is feasible for the commercial production of Zantedeschia plantlets in vitro.

Light-emitting diodes (LEDs) are a potential alternative to tubular fluorescent lamps (TFLs) due to their low forward current, small in size, wavelength specific, solid-state construction, low degradation and long life (Brown et al., 1995; Bula et al., 1991; Fang and Jao, 2000). A light source for young plants production using blue (B) and red (R) LEDs was developed with the capabilities of altering blue and/or red light intensity, R to B ratio, frequency and duty ratio of light (Jao and Fang, 2003). Using such a device, authors found that continuous light from LEDs and TFLs had no significant difference to the growth of potato (Solanum tuberosum L. 'Kennebec') plantlets in vitro. However, intermittent or pulse light promotes the growth of potato plantlets significantly (Jao and Fang, 2004a). Concurrent red and blue light enhances growth of potato plantlets compare with alternating red and blue light under the same daily light integral $(5.53 \text{ mol}\cdot\text{m}^{-2}\cdot\text{d}^{-1})$ and 16/8-h photoperiod (Jao and Fang, 2004b).

Plant growth and development are affected by light intensity, light quality, duration, and photoperiod (Taiz and Zeiger, 1991). Growth, morphology, and differentiation of in vitro

Professor

plantlets are also affected by light quality (Econmou and Read, 1987). Red light was shown significantly enhance stem elongation of Pelargonium plantlets in vitro while blue light inhibited shoot length (Appelgren, 1991). Aksenova et al. (1994) found that in vitro potato plantlets produced longer stems and higher root to shoot ratio in red than in blue. By using TFLs as the main, and red and far-red (FR) LEDs as the supplemental light source, Iwanami et al. (1992) found that the growth of potato plantlets in vitro can be promoted even when the relative proportion of supplemental light was very small. By using red LEDs as the main and using TFLs as the supplemental light source, Miyashita et al. (1994) found that the morphology rather than the growth (dry weight and leaf area) of potato plantlets (Solanum tuberosum L. 'Benimaru') was affected under same PPF.

Zantedeschia are commercial available as cut flowers and potted plants. Tissue-culturederived plantlets are an alternative planting material for tuber or cut flowers production in calla lily (Clemens and Welsh, 1993). However, little information is available on how light quality influences the growth of Zantedeschia plantlets in vitro. The focus of this study was to compare effects of light generated by red and blue light-emitting diodes and TFLs (control group) on the photomixotrophic growth of Zantedeschia plantlets during in vitro stage and later tuber formation after transplant into greenhouse under the same PPF and photoperiod.

Materials and Methods

Plant materials and culture conditions. In vitro plantlets of calla lily (Zantedeschia jucunda 'Black Magic') at final stage were provided by Seed Improvement and Propagation Station (Hsinshe, Taichung, Taiwan) and cultured in 10 ×10-cm, 785-mL cylindrical poly vinyl chloride (PVC) containers (Yu-Ping Corp., Taiwan). Each vessel contained 20 plantlets. In total, 5 treatments and 100 samples were cultured. Vessels were incubated in a controlled environment at 25 \pm 2 °C and 50% \pm 10% RH. Each treatment was arranged in a completely randomized design of 10 individual samples. In total, 50 samples were used for comparison during in vitro stage. Another 50 samples were transplanted and grown using rockwool (Groden, Denmark) culture with drip lines (Netafim, Isreal) in a pad and fan greenhouse for tuber formation. After 6 months, tuber size (shortest length of diameter) was recorded for each sample. Three size groups were used including <3 cm, 3 to 5 cm, and >5 cm. Tubers >3 cm are considered marketable. Percentage of each group and marketable size were calculated. Each treatment was conducted three times.

Light treatments. Cultures of in vitro plantlets were illuminated using red and blue LEDs powered by self-designed driver and TFL light. Photoperiod was 16/8 h (day/night) and photosynthetic photon flux (PPF) was $80 \pm 5 \mu mol \cdot m^{-2} \cdot s^{-1}$. PPF was measured (LI-1800; LI-COR Inc.) when the culture bench was empty. The LED lighting system used was described previously by Jao and Fang (2003, 2004a, 2004b).

Totally, five different light treatments were used in this study (Table 1). Treatment 1 was the control group using TFL. Treatments 2 to 5 used LED as light source. Treatments 2, 3, and 5 used red LED alone with different frequency and duty ratio. Treatment 2 used AC power to light

Table 1. Summary of five treatments.

Treatment	Red light PPF	Blue light PPF	Frequency	Duty ratio	Driver
code	$(\mu mol \cdot m^{-2} \cdot s^{-1})$	$(\mu mol \cdot m^{-2} \cdot s^{-1})$	(Hz)	(%)	used?
1 ^z	48	32	60	50	Ν
2 ^y	80	0	60	50	Ν
3 ^x	80	0	N/A	100	Y
4 ^w	48	32	N/A	100	Y
5 ^v	80	0	60	50	Y

^zControl group using TFL light.

^yRed LED light alone using AC power with 60-Hz frequency and 50% duty ratio.

"Red plus blue LED light were used and powered by driver with R to B ratio equal to 1.5.

vRed LED light alone using DC power from driver with 60-Hz frequency and 50% duty ratio.

Received for publication 5 Aug. 2004. Accepted for publication 26 Sept. 2004.

¹Assistant professor. Corresponding author: e-mail rueychi.jao@msa.hinet.net. ²PhD candidate.

³Professor

^{*}Red LED light alone using DC power from driver with 100% duty ratio.

Table 2. Plant height, chlorophyll content, shoot and root dry and fresh weights per plantlet, percentage of dry matter, and shoot and root dry weight ratio (S to R ratio) on a dry weight basis of the *Zantedeschia* plantlets in vitro 28 d after treatments.

	Plant ht	Chlorophyll content	Fresh wt (g/plantlet)		Dry wt (g/plantlet)			Dry matter	S to R	
Treatment	(cm)	(SPAD)	Shoot	Root	Total	Shoot	Root	Total	(%)	ratio
1	6.3 c ^z	29.9 a	0.289 c	0.426 a	0.715 b	0.023 a	0.043 a	0.066 a	9.18	0.53
2	9.8 a	20.7 c	0.433 a	0.355 b	0.788 a	0.024 a	0.029 b	0.053 b	6.67	0.81
3	10.5 a	19.9 c	0.391 b	0.323 c	0.714 b	0.019 b	0.025 b	0.044 b	6.12	0.79
4	8.5 b	24.2 b	0.312 c	0.303 d	0.614 c	0.018 b	0.027 b	0.045 b	7.29	0.64
5	9.9 a	20.4 c	0.301 c	0.323 c	0.624 c	0.017 b	0.028 b	0.045 b	7.15	0.59

^zMeans within a column followed by the same letters are not significantly different at the 5% level of Duncan's multiple range test.

60 red LEDs connected in series. Therefore, light was emitted naturally with 60-Hz frequency and 50% duty ratio subject to Taiwan's AC system. Treatment 3 used DC power generated from driver with 100% duty ratio. Treatment 5 also used DC power to generate 60 Hz frequency and 50% duty ratio. For treatment 4, blue LED was added and the R to B ratio was maintained at 1.5, the same as for TFL.

Experimental design and statistical analysis. Plant height, chlorophyll content, fresh and dry weight of shoots and roots were measured for each treatment of 50 samples 28 d after planting. During each sampling, chlorophyll content of the youngest fully expanded leaf was measured using portable chlorophyll meter (Minolta SPAD 502; Spectrum Technologies). Percentage of dry matter and shoot (S) to root ratio (R), (S/R, calculated using dry weight) were also calculated. Data were analyzed using Duncan's multiple range test and orthogonal contrasts using SAS (SAS Inst., Cary, N.C.).

Results and Discussion

Table 2 showed that providing the same PPF and photoperiod with red + blue light (Treatments 1 and 4, R/B = 1.5) or red light alone (Treatments 2, 3, and 5) caused the differences in Zantedeschia plantlet height, chlorophyll content and total dry weight during 4 weeks of cultivation. TFL treatment had the highest value on dry weight and chlorophyll content and lower plant height than LED treatments. Contrast treatment 1 with others (Tables 4, 5 and 6) also showed the same results. However, within LED treatments, there were no significant difference in dry weight but differences in chlorophyll content and plant height. When blue light was added, shorter plant height and higher chlorophyll content can be found. Orthogonal contrasts also revealed that there were no significant differences on dry weight between LED treatments (Table 6, contrast treatment 4 with 2, 3, and 5) but in plant height (Table 4, contrast treatment 4 with 2, 3, and 5) and chlorophyll content (Table 5, contrast treatment 4 with 2, 3, and 5). Shorter plant height and higher chlorophyll content can be found in red + blue light cases. It showed that blue light may be involved in plant height and chlorophyll development control. In treatments 1 and 4, R to B ratios were the same, however, TFL treatment derived shortest plant height, highest chlorophyll content and highest dry weight. Orthogonal contrast treatment 1 with 4 (Tables 4, 5, and 6) revealed same result: not only blue light, but also other wavebands of light are involved in this control.

Table 3. Average tuber size in smallest diameter, size distribution and percentage of marketable tuber size in each treatment after 6 months transplanting in greenhouse.

	Tuber	Size	Size	Size	Marketable
	size	<3 cm	3–5 cm	>5 cm	size ^z
Treatment	(cm)	(%)	(%)	(%)	(%)
1	4.2 a ^y	0.0	93.3	6.7	100.0
2	4.1 a	0.0	96.5	3.5	100.0
3	4.0 a	7.1	85.8	7.1	92.9
4	4.5 a	3.7	66.7	29.6	96.3
5	3.9 a	7.8	79.3	12.9	92.2

^zMarketable size means the smallest diameter of the tuber is >3 cm.

^yMeans within a column followed by the same letters are not significantly different at the 5% level of Duncan's multiple range test.

Table 4. Analysis of variance summary for the plant height of *Zantedeschia* plantlets. Data were analyzed using procedures for a completely randomized design with treatment comparisons made using orthogonal contrasts.

Source	df	Sums of squares	Mean square	F value
Treatment	5	52.192	10.439	3.21**
Error	54	175.369	3.247	
Corrected total	59	227.561		
Contrasts				
Treatment 1 vs. others ^z	1	27.421	27.421	8.44**
Treatment 2 vs. 3 ^y	1	3.872	3.872	1.19 ^{NS}
Treatment 4 vs. 2, 3 and 5 ^x	1	13.624	13.624	4.20**
Treatment 1 vs. 5 ^w	1	9.112	9.112	2.81 ^{NS}
Treatment 1 vs. 4 ^v	1	2.964	2.964	4.91**
Treatment 1 vs. 2 ^u	1	16.381	16.381	5.04**

^zTreatment of TFL light alone vs. all other treatments.

^yTreatment of red light alone using AC power vs. treatment using DC power.

^xTreatment of red + blue light vs. red light alone using LED.

"Treatment of 60 Hz using TFL vs. 60 Hz using LED.

^vTreatment of R/B = 1.5 using TFL vs. using LED.

"Treatment of TFL vs. red LED light alone using AC power.

^{NS,**}Nonsignificant or significant contrast at the 0.05 level.

Percentage of dry matter and S to R ratio of plantlets were different within treatments (Table 2). Treatment 1 had the highest dry matter content (about 9%), followed by treatments 4 and 5 (about 7%) and treatments 2 and 3 (about 6%) were the worse. Treatment 2 had the highest S to R ratio and treatment 1 had the lowest. It seemed that TFL treatment produced more root parts than red light alone using LEDs. Besides red light, others bands of light are needed for root parts development.

In the cases of providing red light alone (Treatments 2, 3, and 5), there were no significant differences on plant height, chlorophyll content, dry weight (Table 2) and tuber size (Table 3) no matter how the red light were provided from LEDs. Same results were also revealed on contrast 2 in Tables 4 to 6. The only difference between treatments 2 and 3 was the driving power. Since one red LED consumed 2 v. In total, 60 LEDs were connected in series and powered by AC power with 60 Hz frequency. The main advantage of using AC powered red LEDs was cost down. Costs on AC/DC converter and frequency generator, etc can be removed. Further study is needed to test on the lifetime of LEDs using AC power. Contrast treatment 1 with 5 and treatment 1 with 4 (Tables 5 and 6) showed that there were significant differences between TFL, treatment 2 and treatment 5 in chlorophyll content and dry weight. All treatments had the same frequency of 60 Hz. Treatments 2 and 5 had no differences in plant height, chlorophyll content and dry weight (Table 2). However, differences showed between red LED light alone and TFL. It seemed that not frequency but light quality caused these results.

Results showed that no matter what kind of light provided (TFL or LEDs) or how the current was provided to LEDs (AC or DC), similar tuber size can be produced at the end of growth (first column in Table 3). The influence of light on crops during the in vitro stage does not last to the end of the tuber formation stage. Percentage of marketable size (>3 cm) tuber was >90% in all treatments (Table 3).

In conclusion, blue light was one factor on plant height and chlorophyll development control for *Zantedeschia* plantlets in vitro.

Table 5. Analysis of variance summary for the chlorophyll content of *Zantedeschia* plantlets. Data were analyzed using procedures for a completely randomized design with treatment comparisons made using orthogonal contrasts.

Source	df	Sums of squares	Mean square	F value	
Treatment	5	861.274	172.255	11.27**	
Error	54	825.552	15.288		
Corrected total	59	1686.826			
Contrasts					
Treatment 1 vs. others ^z	1	745.133	745.133	48.74**	
Treatment 2 vs. 3 ^y	1	28.322	28.322	1.85 ^{NS}	
Treatment 4 vs. 2, 3 and 5 ^x	1	77.875	77.875	15.09**	
Treatment 1 vs. 5 ^w	1	587.528	587.528	38.43**	
Treatment 1 vs. 4 ^v	1	242.208	242.208	15.84**	
Treatment 1 vs. 2 ^u	1	389.845	389.845	25.50**	

^zTreatment of TFL light alone vs. all other treatments.

yTreatment of red light alone using AC power vs. treatment using DC power.

^xTreatment of red + blue light vs. red light alone using LED.

"Treatment of 60 Hz using TFL vs. 60 Hz using LED.

vTreatment of R/B=1.5 using TFL vs. using LED.

"Treatment of TFL vs. red LED light alone using AC power.

^{NS,**}Nonsignificant or significant contrast at the 0.05 level.

Table 6. Analysis of variance summary for the dry weight of *Zantedeschia* plantlets. Data were analyzed using procedures for a completely randomized design with treatment comparisons made using orthogonal contrasts.

Source	df	Sums of squares	Mean square	F value
Treatment	5	0.00575	0.001151	6.47**
Error	54	0.00960	0.000177	
Corrected total	59	0.01536		
Contrasts				
Treatment 1 vs. others ^z	1	0.005150	0.005150	28.94**
Treatment 2 vs. 3 ^y	1	0.000055	0.000055	0.31 ^{NS}
Treatment 4 vs. 2, 3 and 5^x	1	0.000016	0.000016	0.09 ^{NS}
Treatment 1 vs. 5 ^w	1	0.004936	0.004936	27.74**
Treatment 1 vs. 4 ^v	1	0.002813	0.002813	15.81**
Treatment 1 vs. 2 ^u	1	0.002215	0.002215	12.45**

^uTreatment of TFL light alone vs. all other treatments.

"Treatment of red light alone using AC power vs. treatment using DC power.

"Treatment of red + blue light vs. red light alone using LED.

^xTreatment of 60 Hz using TFL vs. 60 Hz using LED.

yTreatment of R/B=1.5 using TFL vs. using LED.

^zTreatment of TFL vs. red LED light alone using AC power.

^{NS,**}Nonsignificant or significant contrast at the 0.05 level.

At present, the cost of blue LEDs can be 10 times more than that of red LEDs. Results of this study showed that growth of *Zantedeschia* plantlets in vitro using only AC powered red LEDs is feasible. No significant difference can be found after 6 months of tuber formation growth in greenhouse.

Literature Cited

- Appelgren, M. 1991. Effects of light quality on stem elongation of *Pelargonium* in vitro. Scientia Hort. 45:345–351.
- Aksenova, N.P., T. N. Konstantinova, L.I. Sergeeva, I. Machackova, and S.A. Golyanovskaya. 1994. Morphogenesis of potato plants in vitro. I. Effect of light quality and hormones. J. Plant Growth Regulat. 13:143–146.
- Brown, C.S., A.C. Schuerger, and J.C. Sager. 1995. Growth and photomorphogenesis of pepper plants under red light-emitting diodes with supplemental blue or far-red lighting. J. Amer. Soc. Hort. Sci. 120:808–813.
- Bula, R.J., R.C. Morrow, T.W. Tibbitts, D.J. Barta, R.W. Ignatus, and T.S. Martin. 1991. Lightemitting diodes as a radiation source for plants. HortScience. 26:203–205.
- Clemens, J. and T.E. Welsh. 1993. An overview of the New Zealand calla industry, research directions and year-round tuber production. Acta Hort. 337:161–166.

Economou, A.S. and P.E. Read. 1987. Light treat-

ments to improve efficiency of in vitro propagation system. HortScience 22(5):751–754.

- Fang, W. and R.C. Jao. 2000. A review on artificial lighting of tissue cultures and transplants, p. 108–113. In: C. Kubota and C. Chun (eds.). Transplant production in the 21st century. Kluwer Academic Publishers, The Netherlands.
- Iwanami, Y., T. Kozai, Y. Kitaya and S. Kino. 1992. Effects of supplemental red and far-red lighting using light emitting diode on stem elongation and growth of potato plantlets in vitro. p. 183. In: Abstr. Intl. Symp. Transplant Production Sytems. 21–26 July 1992. Yokohama, Japan.
- Jao, R.C. and W. Fang. 2003. An adjustable light source for photo-phyto related research and young plant production. Appl. Eng. Agr. 19(5):601–608.
- Jao, R.C. and W. Fang. 2004a. Effects of frequency and duty ratio on the growth of potato plantlets in vitro using light emitting diodes. HortScience 39(2):375–379.
- Jao, R.C. and W. Fang. 2004b. Growth of potato plantlets in vitro is different when provided concurrent versus alternating red and blue light photoperiods. HortScience 39(2):380–382.
- Miyashita, Y., T. Kimura, Y. Kitaya, and T. Kozai. 1994. Effects of red light on the growth and morphology of potato plantlets in vitro: Using light emitting diodes (LEDs) as light source for micropropagation. Acta Hort. 418:169–173.
- Taiz, L. and E. Zeiger. 1991. Plant physiology, p. 179–264. 1st ed. Benjamin/Cummings Publ. Co., New York.